



THE UNIVERSITY *of* EDINBURGH

Edinburgh Research Explorer

The contribution of microbially-produced nanoparticles to sustainable development goals

Citation for published version:

Cueva, M & Horsfall, L 2017, 'The contribution of microbially-produced nanoparticles to sustainable development goals', *Microbial biotechnology*. <https://doi.org/10.1111/1751-7915.12788>

Digital Object Identifier (DOI):

[10.1111/1751-7915.12788](https://doi.org/10.1111/1751-7915.12788)

Link:

[Link to publication record in Edinburgh Research Explorer](#)

Document Version:

Publisher's PDF, also known as Version of record

Published In:

Microbial biotechnology

Publisher Rights Statement:

© 2017 The Authors. Microbial Biotechnology published by John Wiley & Sons Ltd and Society for Applied Microbiology. This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

General rights

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy

The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.



The contribution of microbially produced nanoparticles to sustainable development goals

Miguel E. Cueva and Louise E. Horsfall* 

SynthSys, CSEC and School of Biological Sciences,
University of Edinburgh, Edinburgh, UK.

Summary

Nanoparticles (NPs), particles having one or more dimensions below 100 nm, are currently being synthesized through chemical and physical methods on an industrial scale. However, these methods for the synthesis of NPs do not fit with sustainable development goals. NP synthesis, through chemical and physical methods, requires high temperatures and/or pressures resulting in high energy consumption and the generation of large amounts of waste. In recent years, research into the synthesis of NPs has shifted to more green and biological methods, often using microorganisms. A biological approach has many advantages over chemical and physical methods. Reactions are catalysed in aqueous solutions at standard temperature and pressure (cost effective and low energy syntheses). This method does not require solvents or harmful chemicals, making NP biosynthesis a greener and more eco-friendly method. Furthermore, NP synthesis by microbes does not require the use of pure starting materials; thus it can simultaneously be used for the bioremediation of contaminated water, land and waste, and the biosynthesis of NPs. Therefore the biosynthesis of NPs contributes to the sustainable development goals, while the alternative physical and chemical methods exclusively utilize scarce and expensive resources for NP synthesis.

Nanoparticles (NPs), particles having one or more dimensions below 100 nm, are currently being synthesized through chemical and physical methods on an industrial scale. However, these methods for the

synthesis of NPs do not fit with sustainable development goals. NP synthesis, through chemical and physical methods, requires high temperatures and/or pressures resulting in high energy consumption and the generation of large amounts of waste. In recent years, research into the synthesis of NPs has shifted to more green and biological methods, often using microorganisms. A biological approach has many advantages over chemical and physical methods. Reactions are catalysed in aqueous solutions at standard temperature and pressure (cost-effective and low-energy syntheses). This method does not require solvents or harmful chemicals, making NP biosynthesis a greener and more eco-friendly method. Furthermore, NP synthesis by microbes does not require the use of pure starting materials; thus, it can simultaneously be used for the bioremediation of contaminated water, land and waste and the biosynthesis of NPs (Pollman *et al.*, 2006; Macaskie *et al.*, 2010). Therefore, the biosynthesis of NPs contributes to the sustainable development goals, while the alternative physical and chemical methods exclusively utilize scarce and expensive resources for NP synthesis.

Biogenic metal nanoparticles are produced by microorganisms utilizing their natural metal resistance and metabolic pathways, through either an intracellular or an extracellular mechanism. The intracellular mechanism involves ion transportation into the microbial cell where they are then reduced to their elemental form through electrostatic and enzymatic interactions, forming NPs that can be held within the cell. The extracellular mechanism is mediated either through enzymatic reduction at the cell surface or by secreted molecules that reduce metal ions into their elemental form (Hulkoti and Tara-nath, 2014).

Potential applications of biogenic metal NPs range from various biomedical purposes (e.g. antiviral, antibacterial, antiparasite, medical imaging, drug delivery, cancer treatment and medical diagnostics), through environmental remediation purposes (e.g. contaminant/pollutant degradation and catalytic treatments of aqueous organic compounds), to industrial purposes (e.g. enhance fuel cell performance, catalytic organic synthesis and nanoelectrochemistry) (Schröfel *et al.*, 2014). While research using biogenic nanoparticles is relatively new, researchers are testing them for potential applications in biomedical contexts (anticancer, drug delivery and antimicrobial). For instance, AgNPs produced by

Received 27 June, 2017; accepted 1 July, 2017.

*For correspondence. Email: Louise.Horsfall@ed.ac.uk;
Tel. +44 131 650 5363.

Microbial Biotechnology (2017) 00(00), 000–000
doi:10.1111/1751-7915.12788

Funding Information

Engineering and Physical Sciences Research Council, (Grant / Award Number: EP/N026519/1) Biotechnology and Biological Sciences Research Council, (Grant / Award Number: BB/N002520/1) Consejo Nacional de Ciencia y Tecnología

© 2017 The Authors. *Microbial Biotechnology* published by John Wiley & Sons Ltd and Society for Applied Microbiology.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

Bacillus licheniformis have the potential for use as an antiangiogenic (inhibit blood vessel formation to prevent the systemic spread of cancer cells) (Kalishwaralal *et al.*, 2009), while AgNPs produced by *Fusarium oxysporum* have been incorporated into textile fabrics to inhibit or prevent infections from pathogenic bacteria (Durán *et al.*, 2007). Nanomedicine is a growing scientific field and has tremendous prospects for the improvement of the diagnosis and treatment of human diseases, where biogenic NPs will not only be applied, but their production will also fit within sustainable development goals.

Some examples of microorganisms that are capable of synthesizing NPs are as follows: *Desulfovibrio desulfuricans*, *Cupriavidus metallidurans*, *Bacillus subtilis*, *Escherichia coli*, *Rhodococcus* sp., *Candida glabrata* and *Verticillium* sp. (Park *et al.*, 2015). Currently, the range of metals that are able to be produced in nanoparticle form by microbes include (but is not limited to) As, Ag, Al, Au, Cd, Cr, Cu, As, Pb, Pd and Pt (Pantidos and Horsfall, 2014).

Although microbially produced metal NPs have many potential applications, there are still many drawbacks and problems to be addressed in their synthesis. First, biogenic metal NPs may be produced with a broad morphology and size range. Second, their synthesis may not result in sufficiently high yields and, depending on the microorganism used, it may be slow. Third, metal NPs are sometimes synthesized in unstable forms with reduced catalytic activities, and finally, the use of impure starting materials (e.g. waste) affects its downstream purification. In an effort to address these problems, synthetic biology, nanobiotechnology and genetic engineering are being researched to improve NP synthesis. Once size and morphology are controlled, yield is increased and downstream purification enhanced; the overall production of NPs will be both cost-effective and greener (Edmundson *et al.*, 2014).

Synthetic biology involves the design and engineering of microorganisms to make them capable of performing novel functions for industry, medicine and scientific research. Within the different applications of synthetic biology are those exploring the potential biosynthesis of metal NPs and will utilize its modular genetic engineering approach to enhance NP quality and quantity.

Research with cell surface engineering for the synthesis of metal NPs has been carried out through the heterologous expression of synthetic chelators (polypeptide chelators rich in metal binding amino acids, e.g. EC20), phytochelators and metallothionein (cysteine-rich polypeptide chelators found in plants) and phytochelatin synthase (the enzyme responsible for synthesizing phytochelators from glutathione). An early synthetic biology study successfully synthesized Au, Ag, Fe, Te, CdZn, CdSe, CdTe, ZnSe, CdCs, PrGd, SrGd, SrPr, FeAg,

FeCo, FeMn, CdSeZn, FeCoNi, FeCoMn, CdSeZnTe and AuCdSeZn NPs with a recombinant *E. coli* strain co-expressing phytochelatin synthase and metallothionein (Park *et al.*, 2010). While more recently the *in vivo* synthesis of EuSe NPs was performed using recombinant *E. coli* cells expressing phytochelatin synthase, phytochelators and metallothionein, these EuSe NPs exhibited high fluorescence intensities and strong magnetic properties, and their anticancer properties were demonstrated effectively (Kim *et al.*, 2016).

The overexpression of key proteins and enzymes from the NP synthesis pathway can enhance a microorganism's ability to sequester metals and significantly increase their tolerance to metal concentrations. A recent study produced a recombinant *E. coli* strain containing arsenic resistance genes from a number of sources, and it was then demonstrated that the engineered *E. coli* is able to not only tolerate high levels of arsenic, but is also able to convert the arsenic to an insoluble form, thereby removing significant amounts of arsenic from a solution (Edmundson and Horsfall, 2015).

The use of encapsulins also has promising applications within nanobiotechnology. The ability to engineer cellular compartments and functional nanoarchitectures under environmentally sustainable conditions has been fruitful for the synthesis of monodispersed NPs. A recent study engineered the *Thermotoga maritima* encapsulin to yield a designed compartment for the size-constrained synthesis of Ag NPs. Such NPs exhibited a uniform shape and size distribution as well as long-term stability. When their antimicrobial activity was tested, it proved to be superior to that of silver nitrate and citrate-capped Ag NPs (Giessen and Silver, 2016). Another promising application of nanobiotechnology for the synthesis of monodispersed NPs is through the use of *Dps* proteins, oligomeric cage-like molecules that contain ferroxidase centres with iron oxidation/storage capacity. Research on *Dps* proteins from starved *Listeria innocua* cells, and its mutant (lacking catalytic ferroxidase centre) showed they can produce magnetite (Fe₃O₄) NPs, homogeneous NPs with an average diameter of 3 nm inside both non-mutant and mutant cavities (Ceci *et al.*, 2010). An alternative method for the synthesis of magnetite is through the biomineralization of iron with bacterial organelles called magnetosomes. Mms6, a magnetosome protein produced by the magnetotactic bacterium *Magnetospirillum magneticum* AMB-1, has been shown to form magnetite NPs *in vitro*. Recent work which fused Mms6 onto microcontact-printed assembled monolayer surfaces showed control of the formation and location of magnetite NP production in microscale arrays, thus creating magnetic NP patterns (Bird *et al.*, 2016).

Demands for metal NPs have increased as the number of potential applications has grown. But it is hard to

cheaply mass-produce homogenous metal NPs, and no method for the production of biogenic metal NPs has been commercialized. However, recent studies have proved that the scaled-up production of metal NPs, and therefore its future commercialization, is a real possibility (Moon *et al.*, 2010; Byrne *et al.*, 2015). The production of magnetite nanoparticles by the Fe(III)-reducing bacterium *Geobacter sulfurreducens* has successfully been scaled up from the laboratory to pilot plant. The plant-scale production produced up to 120 g of biomagnetite, with a maintained size distribution between 10 and 15 nm, and unchanged surface reactivity and magnetic properties (Byrne *et al.*, 2015). Another recent study illustrating the potential for the commercialization of biogenic NP synthesis performed scale-up experiments with *Thermoanaerobacter* sp. TOR-39 (anaerobe Fe(III)-reducing bacteria), using a 35 l reactor batch fermentation to obtain a yield of over 1 kg (wet weight) of magnetite in less than a month (Moon *et al.*, 2010). Three key aspects for the mass production of NPs through a bacterial fermentation were elucidated in these studies: first, the rate at which particles are produced; second, the size reproducibility (monodispersed NPs); and finally, the ease of downstream recovery. However, the ultimate determinant as to the success of industrial nanoparticles biosynthesis will be the quality of the microbially produced NPs as compared to those produced chemically (Delay and Frimmel, 2012).

The future of microbially produced NPs is likely to involve an increase in the range of metals that microbes can produce NPs from, perhaps to even include nanoparticles made of metals which may not have a chemically derived counterpart. To achieve the aforementioned, the screening of metal NP synthesis from different microorganisms needs to be conducted. Such an approach identified the CuNPs synthesized by *Morganella morganii*, which are far more stable than their commercial counterpart (Ramanathan *et al.*, 2013). A microbial process to produce such NPs on an industrial scale could hypothetically cost a fraction of what a traditional chemical synthesis would cost. If an adequate design of a large-scale bioreactor configuration for the optimization of NPs synthesis is reached and recovery methods are perfected, the future industrial production of biogenic metal NPs seems like a real possibility. Not only would the mass production of biogenic metal NPs be of importance to the scientific world, but it could potentially benefit our society and the environment, generating wealth through the clean-up of metal-contaminated land, water and waste sites and obtaining a final product (metal NPs) that can be reintroduced into the world's economy.

Conflict of interest

None declared.

References

- Bird, S.M., El-Zubir, O., Rawlings, A.E., Leggett, G.J., and Staniland, S.S. (2016) A novel design strategy for nanoparticles on nanopatterns: interferometric lithographic patterning of Mms6 biotemplated magnetic nanoparticles. *J Mater Chem* **4**: 3948–3955.
- Byrne, J.M., Muhamadali, H., Coker, V.S., Cooper, J., and Lloyd, J.R. (2015) Scale-up of the production of highly reactive biogenic magnetite nanoparticles using *Geobacter sulfurreducens*. *J R Soc Interface* **12**: 20150240.
- Ceci, P., Chiancone, E., Kasyutich, O., Bellapadrona, G., Castelli, L., Fittipaldi, M., *et al.* (2010) Synthesis of iron oxide nanoparticles in *Listeria innocua* Dps (DNA-binding protein from starved cells): a study with the wild-type protein and a catalytic centre mutant. *Chem Eur J* **16**: 709–717.
- Delay, M., and Frimmel, F.H. (2012) Nanoparticles in aquatic systems. *Anal Bioanal Chem* **402**: 583–592.
- Durán, N., Marcato, P.D., De Souza, G.I., Alves, O.L., and Esposito, E. (2007) Antibacterial effect of silver nanoparticles produced by fungal process on textile fabrics and their effluent treatment. *J Biomed Nanotech* **3**: 203–208.
- Edmundson, M.C., and Horsfall, L. (2015) Construction of a modular arsenic-resistance operon in *E. coli* and the production of arsenic nanoparticles. *Front Bioeng Biotechnol* **3**: 201500160.
- Edmundson, M.C., Capenness, M., and Horsfall, L. (2014) Exploring the potential of metallic nanoparticles within synthetic biology. *New Biotechnol* **31**: 572–578.
- Giessen, T.W., and Silver, P.A. (2016) Converting a natural protein compartment into a Nanofactory for the size-constrained synthesis of antimicrobial silver nanoparticles. *ACS Synth Biol* **5**: 1497–1504.
- Hulkoti, N.I., and Taranath, T.C. (2014) Biosynthesis of nanoparticle using microbes – A review. *Colloids Surf B Biointerfaces* **121**: 474–483.
- Kalishwaralal, K., Banumathi, E., SurechBabu, R.K., Venkataraman, D., Jeyaraj, M., Soo, H.E., *et al.* (2009) Silver nanoparticles inhibit VEGF induced cell proliferation and migration in bovine retinal endothelial cells. *Colloids Surf B Biointerfaces* **73**: 51–57.
- Kim, E.B., Seo, J.M., Kim, G.W., Lee, S.Y., and Park, T.J. (2016) In vivo synthesis of europium selenide nanoparticles and related cytotoxicity evaluation of human cells. *Enzyme Microb Technol* **95**: 201–208.
- Macaskie, L.E., Mikeheenko, I.P., Yong, P., Deplanche, K., Murray, A.J., Paterson-Beedle, M., *et al.* (2010) Today's wastes, tomorrow's materials for environmental protection. *Hydromet* **104**: 483–487.
- Moon, J., Rawn, C.J., Rondinone, A.J., Love, L.J., Roh, Y., Everett, S.M., *et al.* (2010) Large-scale production of magnetic nanoparticles using bacterial fermentation. *J Ind Microbiol Biotechnol* **37**: 1023–1031.
- Pantidos, N., and Horsfall, L.E. (2014) Biological synthesis of metallic nanoparticles by bacteria, fungi and plants. *J Nanomed Nanotechnol* **5**: 1000233.
- Park, T.J., Lee, S.Y., Heo, N.S., and Seo, T.S. (2010) In vivo synthesis of diverse metal nanoparticles by

- recombinant *Escherichia coli*. *Angew Chem Int Ed Engl* **49**: 7019–7024.
- Park, T.J., Lee, K.G., and Lee, S.Y. (2015) Advances in microbial biosynthesis of metal nanoparticles. *Appl Microbiol Biotechnol* **100**: 521–534.
- Pollman, K., Raff, J., Merroun, M., Fahmy, K., and Selen-ska-Pobell, S. (2006) Metal binding by bacteria from uranium mining waste piles and its technological applications. *Biotechnol Adv* **24**: 58–68.
- Ramanathan, R., Field, M.R., O'Mullane, A.P., Smooker, P.M., Bhargava, S.K., and Bansal, V. (2013) Aqueous phase synthesis of copper nanoparticles: a link between heavy metal resistance and nanoparticle synthesis ability in bacterial systems. *Nanoscale* **5**: 2300–2306.
- Schröfel, A., Kratošová, G., Šafarik, I., Šafariková, M., Raška, I., and Shor, L. M. (2014) Applications of biosynthesized metallic nanoparticles – A review. *Acta Biomater* **10**: 4023–4042.